Molecular Applications in Zoology

Introduction, sample collection, DNA extraction, PCR and electrophoresis



Syllabus

1. Introduction, DNA extraction, PCR (20.2.2024, lecturer Zuzana Starostová)

2. Next generation sequencing – introduction, short-read and long-read sequencing methods, assemblies (27.2.2023, lecturer Radka Reifová)

3. How to read and make phylogenetic trees (5.3.2024, lecturer Zuzana Starostová)

4. Use of molecular phylogenetics in zoology (12.3.2024, lecturer Zuzana Starostová)

5. Microsatellites and molecular identification of species, individuals and sex (19.3.2024, lecturer Pavel Munclinger)

6. Molecular phylogeography (26.3.2024, lecturer Pavel Hulva)

7. Population structure and paternity studies (2.4.2024, lecturer Pavel Munclinger)

8. Next generation sequencing – applications: whole genome sequencing, exome sequencing, ddRAD sequencing, metagenomics (9.4.2024, lecturer Radka Reifová)

9. Functional genetic variability: From SNP to selection (16.4.2024, lecturer Michal Vinkler)

10. Gene expression – quantitative PCR, expression microarrays, transcriptome sequencing and analysis, epigenomics (23.4.2024, lecturer Radka Reifová)

11. Cytogenetic methods and their application in zoology (30.4.2024, lecturer František Šťáhlavský)

Practicals:

Day 1: database search and basic DNA sequence data handling

Day 2: lab work – molecular method of sex determination in birds –PCR, gel electrophoresis + cytogenetic methods

Exam: written test covering content of the lectures

Literature:

https://zuzanastarostova.webnode.cz/news/molecular-applications-in-zoology/



Molecular techniques and zoology

- since 90s molecular techniques more available- automatization, lower costs
- commercial synthesis of oligonucleotides, sequencing services, user friendly kits

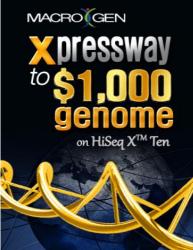
fields like ecology, systematics, ethology, conservation biology...

new methods – new resources - new questions







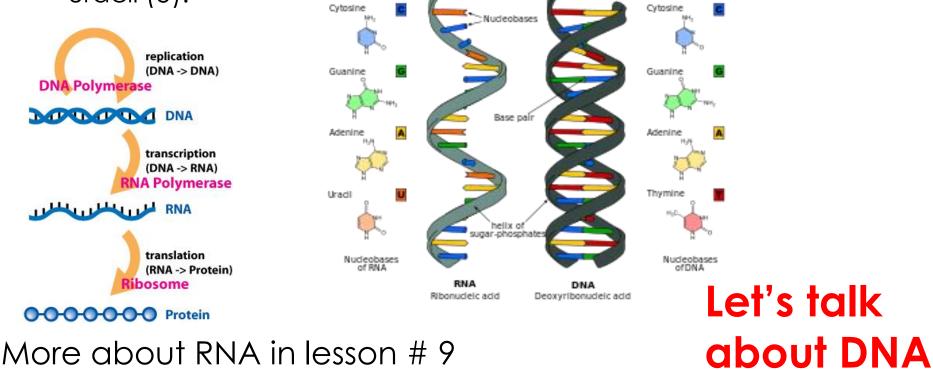


xgenome@macrogen.com

Wikipedia

Nucleic acids

- Nucleic acids are macromolecules that store genetic information and enable protein production
- DNA and RNA molecules composed of strands of nucleotides
- DNA is composed of a phosphate-deoxyribose sugar backbone and the nitrogenous bases adenine (A), guanine (G), cytosine (C), and thymine (T).
- RNA has ribose sugar and the nitrogenous bases A, G, C, and uracil (U).

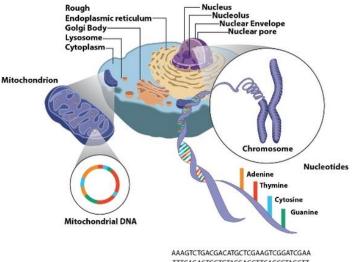


DNA

• is carrying the genetic instructions used in the growth, development, functioning, and reproduction of all known living organisms

Structure:

two polynucleotide strands



- Microsatellite the nitrogenous bases of the two separate polynucleotide strands are bound together, according to base pairing rules (A with T and C with G), with hydrogen bonds to make double-stranded DNA
 - the nitrogenous bases two types: pyrimidines are thymine and cytosine; purines are adenine and guanine
 - two strands of DNA run in opposite directions to each other and are thus antiparallel



Sources of DNA for sampling:

- Destructive sampling tissue (fresh, frozen, dry, preserved in ethanol)
 ideal spleen, muscle- e.g. tongue
- Invasive but non-destructive sampling-buccal smears, fin clips, tail tips (rodents), tissue biopsy, blood sample
- Non-invasive sampling (DNA collected without capturing or usually even touching an animal)- feces, hair, feathers, saliva, shed skin, eggshell
 - DNA yield and quality tend to be lower (higher risk of contamination)



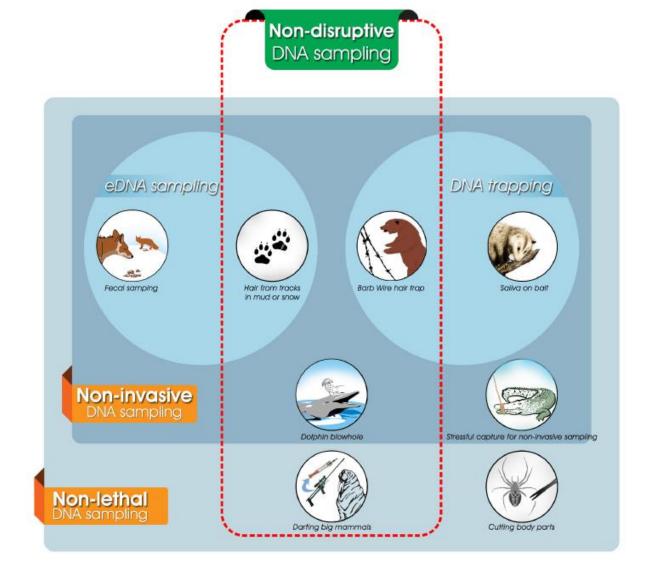


Figure 1. Non-disruptive DNA sampling methods, and their overlaps with noninvasive and non-lethal sampling methods.

Lefort M, Boyer S, Barun A, Emami Khoyi A, Ridden J, Smith VR, Sprague R, Waterhouse BR, Cruickshank RH. (2015) Blood, sweat and tears: non-invasive vs. non-disruptive DNA sampling for experimental biology. PeerJ PrePrints 3:e1580 https://doi.org/10.7287/peerj.preprints.655v3

fur traps





- different for large vs. small species
- effectiveness increased by attractants cats Valeriana officinalis, deer anise





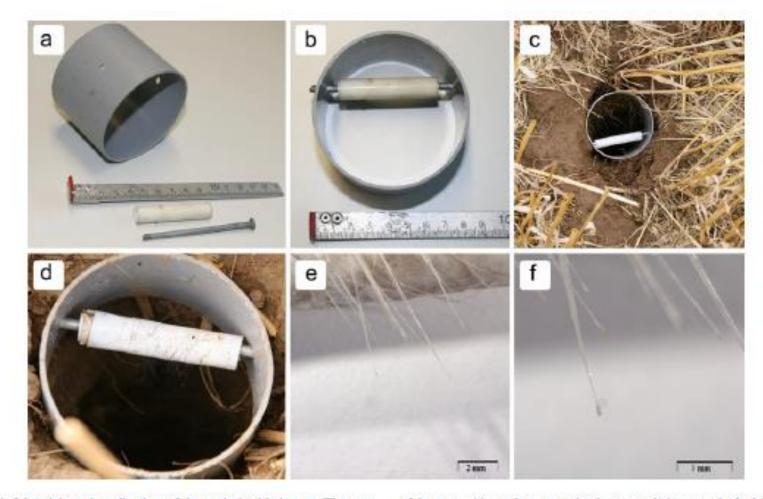


Fig. 1 Materials and application of the optimized hair trap. The trap consists of a PVC pipe with two holes, an iron nail, and a plastic roll (a). The plastic roll is wrapped with double-faced adhesive tape and attached inside the pipe by the nail (b). Traps are placed in entrances



of burrows (c), and as an animal passes, hairs are plucked by the rolling tape (d). Most of plucked hair contained bulbs with a sufficient amount of follicular tissue (e and f)

> Eur J Wildl Res (2011) 57:991-995 DOI 10.1007/s10344-011-0543-9

TECHNICAL NOTES

An optimized hair trap for non-invasive genetic studies of small cryptic mammals

Tobias Erik Reiners · Jorge A. Encarnação · Volkmar Wolters

eDNA

Environmental DNA or **eDNA** is DNA that is collected from a variety of environmental samples (soil, water, snow, air) rather than directly sampled from an individual organism

cave-dwelling amphibian *Proteus anguinus* inhabits subterranean waters of the north-western Balkan Peninsula. Because only fragments of its habitat are accessible to humans, this endangered salamander's exact distribution has been difficult to establish

SCIENTIFIC REPORTS

OPEN Environmental DNA in subterranean biology: range extension and taxonomic implications for *Proteus*

Received: 17 October 2016 Accepted: 20 February 2017 Published: 27 March 2017

Špela Gorički^{1,4} ", David Stanković^{1,2,3,*,*}, Aleš Snoj², Matjaž Kuntner⁴, William R. Jeffery⁵, Peter Trontelj⁶, Miloš Pavićević⁷, Zlatko Grizelj⁸, Magdalena Năpăruș-Aljančić^{1,9} & Gregor Aljančić¹

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Loch Ness Monster may be a giant eel, say scientists

© 5 September 2019

THE LOCH NESS **MONSTER?**

Of the common theories associated with the 1,000 or so sightings of something swimming in the water at Loch Ness, the environmental DNA data obtained suggests at least one theory remains plausible. Eels returned the largest proportion of DNA from the 250 water samples taken throughout Loch Ness.

 \bigtriangledown - <

Typically not gigantic, could an extremely large European eal be the creature people have seen moving "like a torpedo" in the water? The dato abtained suggests this may be possible, although no eal of the size described in some accounts has ever been caught or found.

Infrequent visitors such as seals and possibly sturgeons may account for some sightings, but wakes, standing waves and logs are the basis of most.





1' 2' 3' 8' 9' 10' 11' 12' 5' 6' A LOCH NESS EEL?





CATFISH

OTAGO

Te Whare Wänanga o Otägo NEW ZEALAND

4



Peer ∪

eDNAir: proof of concept that animal DNA can be collected from air sampling

Elizabeth L. Clare, Chloe K. Economou, Chris G. Faulkes, James D. Gilbert, Frances Bennett, Rosie Drinkwater and Joanne E. Littlefair

School of Biological and Chemical Sciences, Queen Mary University of London, London, United Kingdom

ABSTRACT

Environmental DNA (eDNA) is one of the fastest developing tools for species biomonitoring and ecological research. However, despite substantial interest from research, commercial and regulatory sectors, it has remained primarily a tool for aquatic systems with a small amount of work in substances such as soil, snow and rain. Here we demonstrate that eDNA can be collected from air and used to identify mammals. Our proof of concept successfully demonstrated that eDNA sampled from air contained mixed templates which reflect the species known to be present within a confined space and that this material can be accessed using existing sampling methods. We anticipate this demonstration will initiate a much larger research programme in terrestrial airDNA sampling and that this may rapidly advance biomonitoring approaches. Lastly, we outline these and potential related applications we expect to benefit from this development.

Subjects Animal Behavior, Biodiversity, Conservation Biology, Ecology, Zoology Keywords airDNA, eDNA, Biomonitoring, Biodiversity, Terrestrial



More: https://www.the-scientist.com/environmental-dna-can-be-pulled-from-the-air-68645

Clare at al. 2021. eDNAir: proof of concept that animal DNA can be collected from air sampling. PeerJ 9:e11030 https://doi.org/10.7717/peerj.11030

Special challenges:

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DNA Extraction from Dry Museum Beetles without Conferring External Morphological Damage

M. Thomas P. Gilbert^{1,2}*, Wendy Moore³, Linea Melchior^{1,4}, Michael Worobey¹



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The Isolation of Nucleic Acids from Fixed, Paraffin-Embedded Tissues–Which Methods Are Useful When?

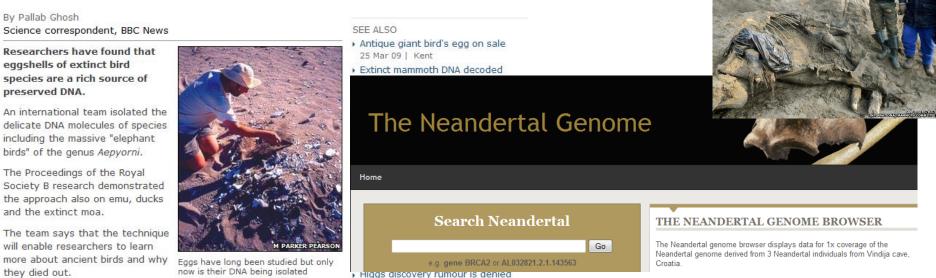
M. Thomas P. Gilbert^{1r}*, Tamara Haselkom¹, Michael Bunce², Juan J. Sanchez³, Sebastian B. Lucas⁴, Laurence D. Jewell⁵, Eric Van Marck⁶, Michael Worobey¹



Fossils as a source of DNA – ancient DNA

- authentic ancient DNA up to several hundred thousand years old samples
- bones, artificially or naturally mummified animal remains, remains in permafrost, fossil eggs, paleofaeces
- extremely high risk of contamination-specialized laboratories
- typically short fragments of DNA, but now also whole genomes (e.g.: mt genomes of mammoth, moas, whole genome of the Neanderthal)

DNA of extinct birds extracted from ancient eggshell





Storing samples for DNA extraction:

- frozen (-20°C usually OK, -80°C for long periods)
- dry
- fixed tissue in 96% ethanol (formaldehyde not ideal, but not impossible)
- samples for RNA isolation can be stored in RNAlater (or -80°C)



Genetic banks:

GLOBAL REGISTRY OF BIODIVERSITY REPOSITORIES



Institutions Institutional/Project Collections Personal Collections Staff Members

Biorepository: Department of Zoology, Charles University in Prague

View Record Edit Record Add Collection Add

Add Staff Member

Institution Information



DNA extraction

- different methods and technologies are available
- all methods involve disruption and lysis of the starting material followed by the removal of proteins and other contaminants and finally recovery of the DNA
- removal of proteins proteinase K
- choice of a method depends on: the required quantity of the DNA, the purity required for downstream applications, and the time and costs

Examples of methods:

- organic extraction phenol/chloroform
- specialized kits usually using binding of the DNA to a solid-phase support





Table 1. Maximum Amounts of Starting Material

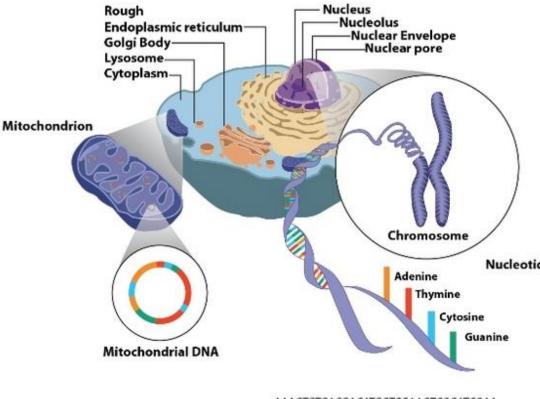
Sample	Amount	
Animal tissue (see Table 3, page 23)	25 mg (spin-column protocols)	6
	20 mg (DNeasy 96 protocols)	
Mammalian blood (see Table 4, page 23)	100 µl	
Bird or fish blood (with nucleated erythrocytes)	10 µl	
Mouse tail	0.6–1.2 cm	
Rat tail	0.6 cm	
Cultured cells	5 x 10 ⁴	Read
Bacteria	2 x 10°	- ACUL





r-to-use DNA

Different genes or genomes = different questions



AAAGTCTGACGACATGCTCGAAGTCGGATCGAA TTTCAGACTGCTGTACGAGCTTCAGCCTAGCTT

Microsatellite

Location of **mtDNA and nuclear DNA in an animal cell**. The mitochondria of an animal cell harbor maternally transmitted mtDNA. Nuclear DNA, which is transmitted by both parents, is found in the nucleus, and contains repeated elements termed microsatellites. © 2012 Nature Education

nuclear DNA

-biparental inheritance in diploid sexually reproducing organisms

-nuclear coding genes- lower mutation rate (in mammals mtDNA ca 10x higher Nucleotides mutation rate)

> -microsatellite loci - mutate much more rapidly than most other types of sequences

> -sex chromosomes - in mammals, the Y chromosome is the paternally inherited most of it does not undergo recombination - useful marker for retracing male lineages

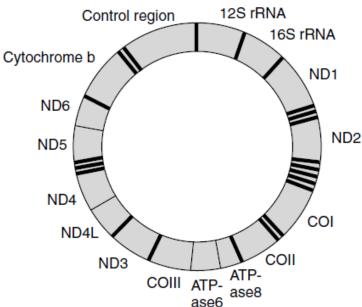
Different genes or genomes = different questions

mitochondrial DNA

-uniparentally inherited-maternal (but exceptions exist)

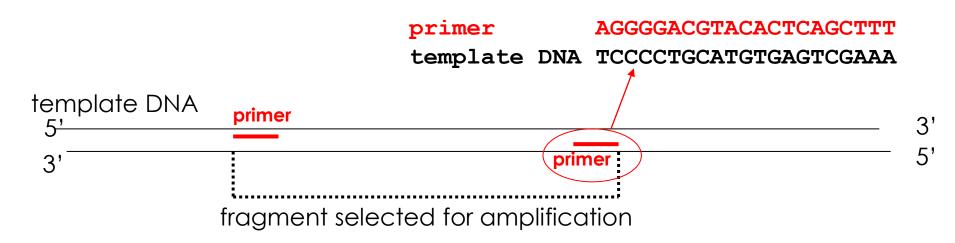
- -mtgenomes relatively small-between 15 000 and 17 000 bp
- -structure, size and arrangement of genes are relatively conserved
- no recombination = offspring will usually have exactly the same mitochondrial genome as its mother Control region
- the mutation rate is relatively high
- non-coding control region (incl. D-loop), evolves rapidly in many taxa
- -small effective population size sensitive to demographic events such as bottlenecks
- -available universal primers

Figure 2.1 Typical gene organization of vertebrate mtDNA. Unlabelled dark bands represent 22 transfer RNAs (tRNAs). Gene abbreviations starting with ND are subunits of NADH dehydrogenase and those starting with CO are subunits of cytochrome *c*. Molecular Ecology, 2nd Edition



Polymerase Chain Reaction = PCR

- method for rapid amplification of DNA sequences selected fragment of DNA
- amplified fragment of DNA is defined by pair of primers
- Primer short DNA fragments containing sequences complementary to the target region



Polymerase Chain Reaction = PCR



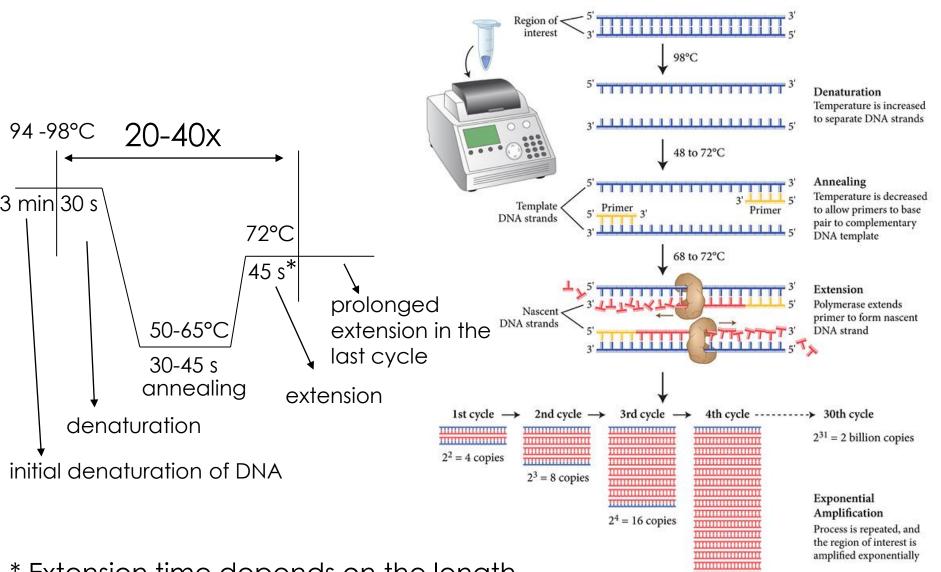
• developed in mid 80s by Kery Mullis

• 1993 – Kery Mullis and Michael Smith awarded the Nobel Prize in Chemistry for their work on PCR

PCR uses repeated cycles of heating and cooling to make many copies of a specific region of DNA

3 steps

- 1. temperature is raised to near boiling causing the double-stranded DNA to separate (denature) into single strands
- 2. temperature is decreased and short DNA sequences (primers) bind (anneal) to complementary matches on the target DNA sequence
- 3. at a slightly higher temperature the enzyme Taq polymerase binds to the primed sequences and adds nucleotides to extend the second strand



* Extension time depends on the length of amplified fragment- nucleotides are incorporated in the rate cca 35 nucleotides per second

 $2^5 = 32$ copies

Polymerase Chain Reaction = PCR

PCR requires:

- two specific oligonucleotide primers
- thermostable DNA polymerase Taq polymerase
- dNTP's
- template DNA
- MgCl₂
- buffer (supplied with the polymerase)
- water

PCR is performed in the thermal cycler (also known as a thermocycler or PCR machine)

PCR usually prepared in small volumes – 20-100 µl





PCR primers

- short oligonucleotides 17-30bp long
- sequences complementary to the target region
- no repetitive sequences
- no sequence complementarity within or between primers secondary structures, primer dimer
- annealing temperature dependent upon primer sequence (~ 50% GC content)
- software for primer design (web tools)

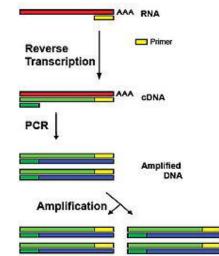
RT-PCR (reverse transcription PCR) and Real-Time PCR

RT-PCR – method to detect RNA expression

- modification of PCR, RNA is template
- qualitatively detect gene expression through creation of complementary DNA (cDNA) transcripts from RNA

Real-Time PCR (also qPCR)

- quantitatively measure the amplification of DNA using fluorescent probes
- specially modified cycler



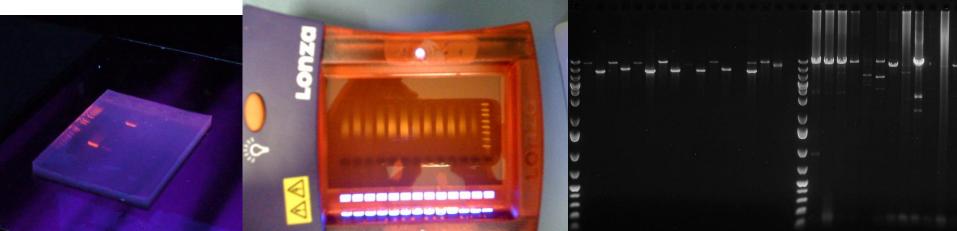


Gel electrophoresis

method for separation and analysis of macromolecules (DNA, RNA, proteins) based on their size and charge electrophoresis = transport by electricity

-**DNA** is negatively charged – fragments of different length are separated by applying an electric field - they move through a gel (usually agarose gel, for more precise separation polyacrylamide gel)

-shorter molecules move faster and migrate farther than longer ones



Gel electrophoresis

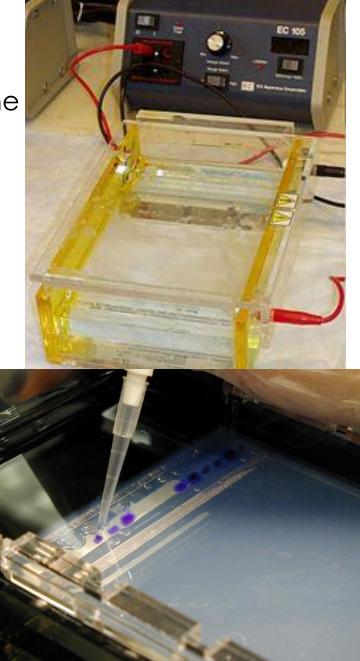
 electric field consists of a negative charge at one end which pushes the molecules through the gel, and a positive charge at the other end that pulls the DNA

- molecules being sorted are dispensed into a well in the gel material

- gel is placed in an electrophoresis chamber, which is then connected to a power source

- when the electric current is applied fragments molecules move

-the different sized molecules form distinct bands on the gel



Gel electrophoresis

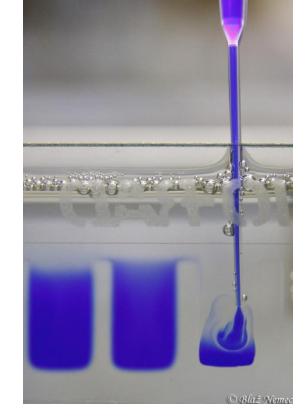
 electrophoresis chamber is filled with buffers
 provide ions that carry a current and maintain stable pH

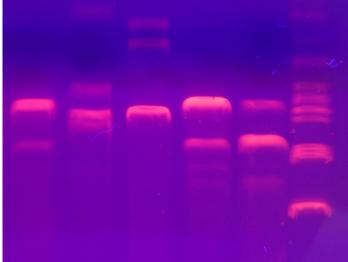
- most common buffers for nucleic acids Tris/Acetate/EDTA (TAE) or Tris/Borate/EDTA (TBE) buffers

- samples are mixed with dye for monitoring

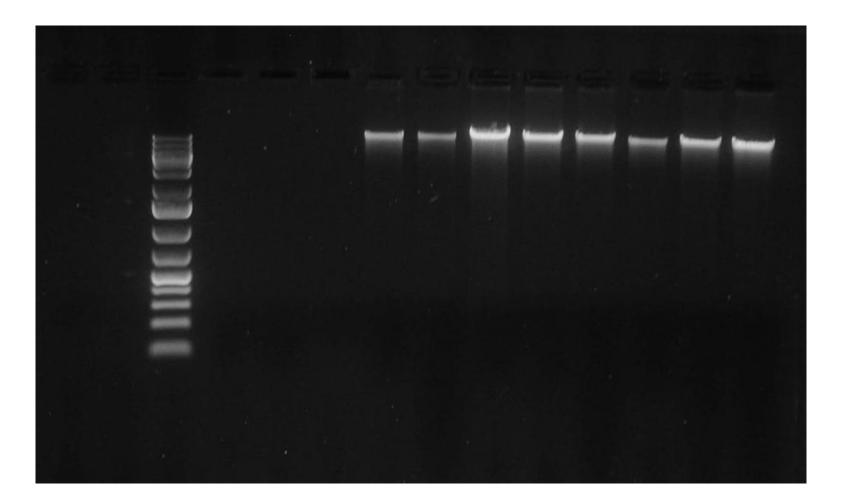
-to visualize DNA fragments - ethidium bromide bind to the DNA - visible under UV (SYBR Green also used)

-size standard (ladder) for estimating size of fragments

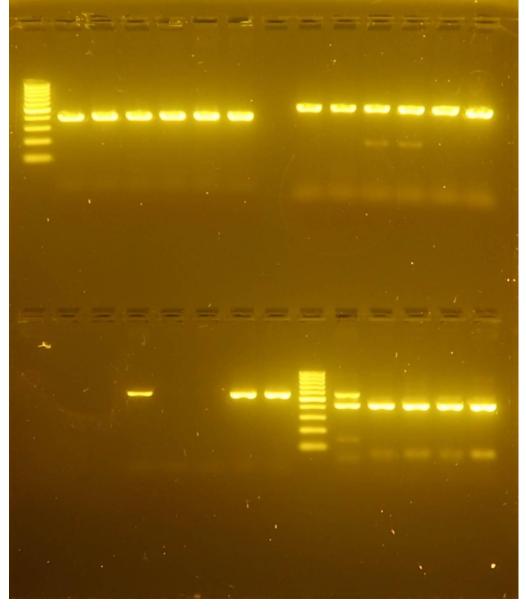




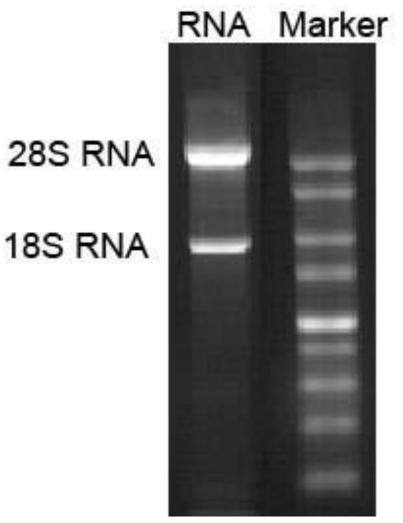
ELFO of total genomic DNA

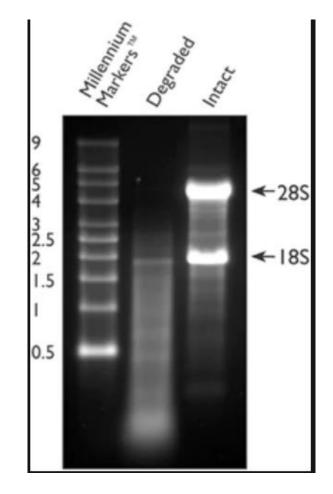


ELFO after PCR



RNA gel electrophoresis





www.labguide.cz

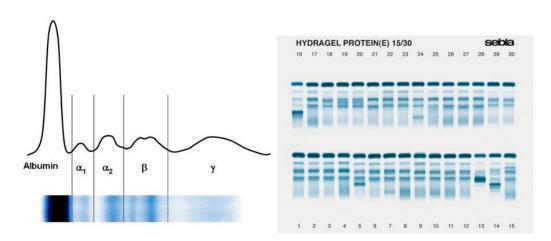
www.thermofisher.com

Protein electrophoresis:

at pH 8-9 proteins- negatively charged = they migrate through the gel while exposed to an electric current (proteins with a negative charge will migrate towards the positively charged anode)

common use: medicine – e.g. electrophoresis of blood serum – information about type of inflammation – acute/chronic, disease diagnostics ...

In case of blood serum – separation into five major fractions by size and electrical charge: serum albumin, alpha-1 globulins, alpha-2 globulins, beta 1 and 2 globulins, and gamma globulins



Characteristic Patterns of Acute-Reaction Proteins Found on Serum Protein Electrophoresis and Associated Conditions or Disorders

Increased albumin

Dehydration

Decreased albumin Chronic cachectic or wasting diseases Chronic infections Hemorrhage, burns, or protein-losing enteropathies Impaired liver function resulting from decreased synthesis of albumin

Malnutrition Nephrotic syndrome Pregnancy

Increased alpha₁ globulins Pregnancy

Decreased alpha₁ globulins Alpha₁-antitrypsin deficiency

Increased alpha₂ globulins Adrenal insufficiency Adrenocorticosteroid therapy Advanced diabetes mellitus Nephrotic syndrome

Decreased alpha₂ globulins Malnutrition Megaloblastic anemia Protein-losing enteropathies Severe liver disease Wilson's disease Increased beta1 or beta2 globulins Biliary cirrhosis Carcinoma (sometimes) Cushing's disease Diabetes mellitus (some cases) Hypothyroidism Iron deficiency anemia Malignant hypertension Nephrosis Polvarteritis nodosa Obstructive jaundice Third-trimester pregnancy Decreased beta₁ or beta₂ globulins Protein malnutrition Increased gamma globulins Amyloidosis Chronic infections (granulomatous diseases) Chronic lymphocytic leukemia Cirrhosis Hodgkin's disease Malignant lymphoma Multiple myeloma Rheumatoid and collagen diseases (connective tissue disorders) Waldenström's macroglobulinemia Decreased gamma globulins Agammaglobulinemia Hypogammaglobulinemia

Protein electrophoresis and zoology:

method: variation in the electrophoretic mobilities of proteins can be used as a measure of nucleotide sequence variation in nuclear DNA

- genetic variability within population, cryptic species, phylogeny reconstruction

samples: tissues, blood, saliva, urine



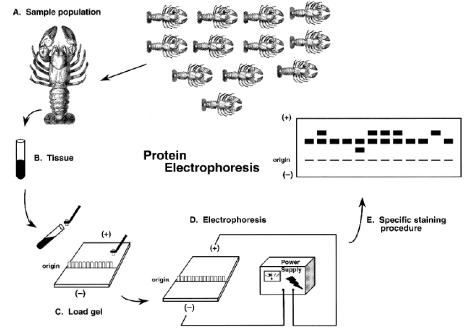


Fig. 8.1 Graphic depicting the steps involved in the protein electrophoresis method. A Drawing samples from a natural population. B Dissecting a tissue sample from an individual to make a tissue extract. C Loading the extract onto a starch gel using filter paper wicks. D Running a cur^rent through the gel to separate the proteins by charge and size. E Adding a specific staining recipe to the sliced gel to visualise specific protein products (see also Plate 29).

Examples of the use of PCR + ELFO methods:

- DNA sequencing – Sanger sequencing – next lecture

- molecular sex determination – more on March 19

- SSCP

PROTOCOL

Single-strand conformation polymorphism (SSCP) for the analysis of genetic variation

Robin B Gasser¹, Min Hu¹, Neil B Chilton², Bronwyn E Campbell¹, Aaron J Jex¹, Domenico Otranto³, Claudia Cafarchia3, Ian Beveridge1 & Xingquan Zhu1,4

¹Department of Veterinary Science, The University of Melbourne, 230 Princes Highway, Werribee, Victoria 3030, Australia. ²Department of Biology, University of Adoptional Stakatchevam, Sakatchevam, Correspondence should be addressed to R.B.G. (robinbg@unimelb.edu.au).

Published online 31 January 2007; doi:10.1038/nprot.2006.485

- after denaturation, single-stranded DNA undergoes 3-dimensional folding = unique conformational state based on its DNA sequence
- difference in "shape" between two singlestranded DNA strands with different sequences can cause them to migrate differently through an electrophoresis gel (number of nucleotides still the same)

use: intraspecific studies: e.g. genotyping to detect homozygous/heterozygous individuals; virology- virus variants

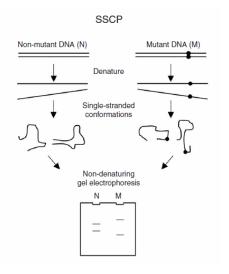
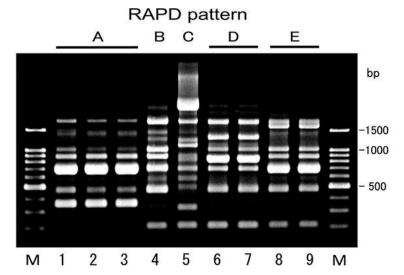


Figure 1 | The principle of PCR-based SSCP analysis. A point mutation (represented by a dot on a DNA strand) leads to the formation of different single-strand conformations of the mutant DNA (M) compared with the nonmutant molecule (N), resulting in differential mobilities in a non-denaturing gel matrix.

RAPD = randomly amplified polymorphic DNA

- does not require any specific knowledge of the DNA sequence of the target organism
- markers are DNA fragments from PCR amplification of random segments of genomic DNA with single primer of arbitrary nucleotide sequence (primers 8-12 nucleotides)
- PCR reaction with one primer, if primers bind to the DNA in reasonable proximity – fragment is amplified, primers bind to the genomic DNA on many positions = fragments of different length – species specific pattern
- Χ
- method sensitive to lab conditions, problems with reproducibility



Use: RAPD is an inexpensive yet powerful typing method for many bacterial species.

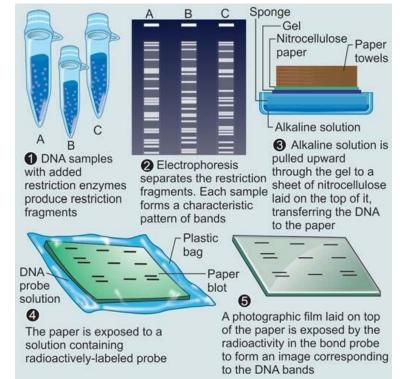
Figure. Random amplified polymorphic DNA (RAPD) patterns of CTX-M-2 β-lactamase-producing Escherichia coli isolated from cattle. Lanes M, 100-bp DNA ladder; lanes 1–9, strains GS528, GS542, GS547, GS553, GS554, GS721, GS733, GS631, and GS671, respectively. Five RAPD patterns, A to E, were produced with RAPD analysis primer 4 (Amersham Pharmacia Biotech, Piscataway, NJ). Shiraki et al. 2004

RFLP – Restriction Fragment Length Polymorphism

restriction enzyme sites - specific sequences of nucleotides (4-8 base pairs in length), which are recognized by restriction enzymes

Method: DNA sample is digested into fragments by one or more restriction enzymes, and the resulting restriction fragments are then separated by gel electrophoresis according to their size, after that transferred to a membrane via the Southern blot procedure. Hybridization of the membrane to a labeled DNA probe then determines the length of the fragments which are complementary to the probe

Use: formerly a tool in genome mapping and genetic disease analysis, basis for early methods of genetic fingerprinting - useful in the identification of samples retrieved from crime scenes, in the determination of paternity, and in the characterization of genetic diversity ...

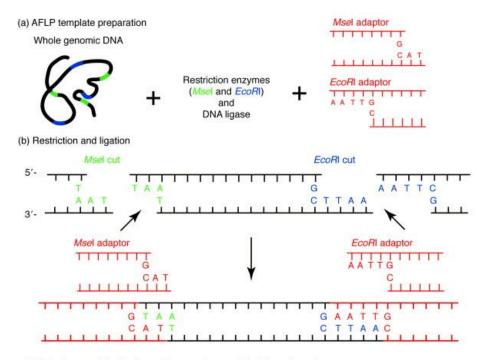


AFLP = amplified fragment length polymorphism

- PCR-based technique that uses selective amplification of a subset of digested DNA fragments to generate and compare unique fingerprints for genomes of interest

- does not require prior information regarding the targeted genome, high reproducibility and sensitivity for detecting polymorphism at the level of DNA sequence

Use: to assess genetic diversity within species or among closely related species, to infer population-level phylogenies and biogeographic patterns, to generate genetic maps - template can be also cDNA – then useful for studying differences in gene expression and searching for target genes



(c) Selective amplification (one of many primer combinations shown)

 Msel primer 1

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